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Please find below and/or attached an Office communication concerning this application or proceeding.

The time period for reply, if any, is set in the attached communication.

Office Action Summary

Application No.

10/673,038

Applicant(s)

GRIMM ET AL.

Examiner

Steven C. Pohnert

Art Unit

1634

Period for Reply -- The MAILING DATE of this communication appears on the cover sheet with the correspondence address --

A SHORTENED STATUTORY PERIOD FOR REPLY IS SET TO EXPIRE 3 MONTH(S) OR THIRTY (30) DAYS, WHICHEVER IS LONGER, FROM THE MAILING DATE OF THIS COMMUNICATION.

- Extensions of time may be available under the provisions of 37 CFR 1.136(a). In no event, however, may a reply be timely filed after SIX (6) MONTHS from the mailing date of this communication.
- If NO period for reply is specified above, the maximum statutory period will apply and will expire SIX (6) MONTHS from the mailing date of this communication.
- Failure to reply within the set or extended period for reply will, by statute, cause the application to become ABANDONED (35 U.S.C. § 133). Any reply received by the Office later than three months after the mailing date of this communication, even if timely filed, may reduce any earned patent term adjustment. See 37 CFR 1.704(b).

Status

- 1) ☒ Responsive to communication(s) filed on 20 August 2008.
- 2a) ☒ This action is **FINAL**. 2b) ☐ This action is non-final.
- 3) ☐ Since this application is in condition for allowance except for formal matters, prosecution as to the merits is closed in accordance with the practice under *Ex parte Quayle*, 1935 C.D. 11, 453 O.G. 213.

Disposition of Claims

- 4) ☒ Claim(s) 1-7 and 10-15 is/are pending in the application.
- 4a) Of the above claim(s) 14 is/are withdrawn from consideration.
- 5) ☐ Claim(s) _____ is/are allowed.
- 6) ☒ Claim(s) 1-7, 10-13 and 15 is/are rejected.
- 7) ☐ Claim(s) _____ is/are objected to.
- 8) ☐ Claim(s) _____ are subject to restriction and/or election requirement.

Application Papers

- 9) ☐ The specification is objected to by the Examiner.
- 10) ☒ The drawing(s) filed on 29 September 2003 is/are: a) ☒ accepted or b) ☐ objected to by the Examiner.
- Applicant may not request that any objection to the drawing(s) be held in abeyance. See 37 CFR 1.85(a).
- Replacement drawing sheet(s) including the correction is required if the drawing(s) is objected to. See 37 CFR 1.121(d).
- 11) ☐ The oath or declaration is objected to by the Examiner. Note the attached Office Action or form PTO-152.

Priority under 35 U.S.C. § 119

- 12) ☐ Acknowledgment is made of a claim for foreign priority under 35 U.S.C. § 119(a)-(d) or (f).
- a) ☐ All b) ☐ Some * c) ☐ None of:
1. ☐ Certified copies of the priority documents have been received.
 2. ☐ Certified copies of the priority documents have been received in Application No. _____.
 3. ☐ Copies of the certified copies of the priority documents have been received in this National Stage application from the International Bureau (PCT Rule 17.2(a)).

* See the attached detailed Office action for a list of the certified copies not received.

Attachment(s)

- 1) ☐ Notice of References Cited (PTO-892)
- 2) ☐ Notice of Draftsperson's Patent Drawing Review (PTO-948)
- 3) ☐ Information Disclosure Statement(s) (PTO/SB08)
Paper No(s)/Mail Date _____
- 4) ☐ Interview Summary (PTO-413)
Paper No(s)/Mail Date _____
- 5) ☐ Notice of Informal Patent Application
- 6) ☐ Other: _____

DETAILED ACTION

Claims Status

This action is in response to the papers filed on 8/20/2008. Currently claims 1-7 and 10-15 are pending.

Claim 14 is withdrawn as directed to a non-elected invention in response to restriction of 5/4/2006.

The claim objection has been withdrawn in view of the arguments that reference of Table 1 is the most practical way to write the claim.

The 112-1st paragraph Written Description rejection has been withdrawn in view of the amendment.

This action is final.

Election/Restrictions

1. This application contains claim 14 drawn to an invention nonelected with traverse in the reply filed on 6/5/2005. A complete reply to the final rejection must include cancellation of nonelected claims or other appropriate action (37 CFR 1.144) See MPEP § 821.01.

Claim Rejections - 35 USC § 112-Maintained

2. The following is a quotation of the first paragraph of 35 U.S.C. 112:

The specification shall contain a written description of the invention, and of the manner and process of making and using it, in such full, clear, concise, and exact terms as to enable any person skilled in the art to which it pertains, or with which it is most nearly connected, to make and use the same and shall set forth the best mode contemplated by the inventor of carrying out his invention.

3. Claims 1-7 and 10-13 and 15 are rejected under 35 U.S.C. 112, first paragraph, because the specification, while being enabling for a method for detecting the presence of a TEM beta-lactam resistant micro-organism in a bacterial sample, said method comprising (i) obtaining a bacterial sample; (ii) isolating DNA contained in the sample; (iii) contacting the DNA of the sample with a micro-array, harboring on predetermined locations thereon different sets of capture probes, under conditions allowing hybridization of complementary strands, (a) wherein each representative of a set of capture probes comprises the sequence R1-(X)-R2, wherein X consists of the nucleotide triplets of amino acid positions 2, 3, 4, 19, 37, 40, 49, 67, 78, 82, 90, 102, 113, 122, 125, 127, 143, 151, 161, 162, 163, 180, 182, 194, 202, 216, 235, 236, 237, 241, 258, 261, 264, 271, 272, 276, and 285 of the bla-TEM-1 gene and R1 consists of 25 consecutive nucleotides immediately 5' to X in the TEM1 and R2 consists of 25 consecutive nucleotides immediately 3' of X, (b) wherein the different sets of capture probes are selected such that an adjacent set starts at a given position 3n of nucleotides downstream from the first set of capture probes, so that the nucleotide sequence of the beta-lactamase gene is covered over a desired range, and (iv) determining if occurrence hybridization is indicative of (i) the presence of a TEM beta-lactam resistant micro-organism, does not reasonably provide enablement for a method of detecting the presence of "any" beta lactam resistant micro-organism in "any" biological sample and determining the particular polymorphism present in the TEM beta-lactam gene. The specification does not enable any person skilled in the art to which it pertains, or with

which it is most nearly connected, to make and use the invention commensurate in scope with these claims.

There are many factors to be considered when determining whether there is sufficient evidence to support that a disclosure does not satisfy the enablement requirement and whether any necessary experimentation is undue. These factors have been described by the court in *re Wands*, 8 USPQ2d 1400 (CA FC 1988). *Wands* states at page 1404,

"Factors to be considered in determining whether a disclosure would require undue experimentation have been summarized by the board in the *Ex parte Forman*. They include (1) the quantity of experimentation necessary, (2) the amount of direction or guidance presented, (3) the presence of working examples, (4) the nature of the invention, (5) the state of the prior art, (6) the relative skill of those in the art, (7) the predictability or unpredictability of the art, and (8) the breadth of the claims."

The nature of the invention and the breadth of the claims:

The claims broadly encompass a method of detecting the presence of a beta-lactam resistant micro-organism in a biological sample comprising: obtaining a biological sample, optionally isolating DNA from the sample, contacting the DNA of the sample with a microarray harboring at predetermined locations different sets of capture probes, under conditions that allow for hybridization of complementary strands, wherein each representative set of probes comprises a sequence R1-(X)-R2, wherein each of said sets consists of at least two representatives and each representative of a set of

capture probes consists of the sequence R1-(X)-R 2, which sequence represents a selected part of the sequence of a beta- lactamase gene among the TEM beta-lactamases exhibiting Extended Spectrum (ESBL) or Inhibitor Resistant (IRT) phenotype, wherein X consists of a nucleotide triplet identified in of table 1 as either a wild type or mutant sequence of TEM beta-lactamase, and wherein R1 consists of 3 to 20 consecutive nucleotides immediately 5' to X in the TEM beta-lactamase and R2 consists of 3 to 20 consecutive nucleotides immediately 3' to X in the TEM beta-lactamase and determining whether hybridization occurs and at which position wherein the occurrence and location of hybridization on the array is indicative of the presence of bet-lactam resistance and the genotype.

The claims thus broadly encompass detection of beta lactam resistance by the use of a microarray with probes comprising "any" triplet of nucleotides in table 1 with "any" length of nucleotides on either side of the triplet that are specific to "any" TEM beta lactamase gene.

The claims further encompass the detection of beta lactam resistance by the presence of "any" hybridization to the array.

Further the claims encompass identification of the particular polymorphism for the location and presence of hybridization.

Claim 15 is drawn to a method of detecting the presence of a beta-lactam resistance microorganism in a biological sample and simultaneously determining the genotype of beta-lactam resistance comprising: obtaining a biological sample; optionally isolating DNA from the sample; contacting the DNA of the sample with a microarray

having the nucleic acid sequence of SEQ ID NO 5 to 45 under conditions allowing for hybridization; and determining whether hybridization occurs and if hybridization occurs the occurrence and location of hybridization is indicative of the presence of beta lactam resistant microorganism and a particular polymorphism of the TEM beta lactamase gene.

Thus claim 15 like claim 1 is drawn to the presence of hybridization of probes as claimed be indicative of beta-lactam resistant microorganisms.

The amount of direction or guidance and the Presence and absence of working examples.

The specification teaches over 340 different beta-lactamase sequences have been identified (0002). The specification teaches that antibiotic resistance in ESBL- and IRT-enzymes are mainly due to 37 amino acid substitutions in 7 amino acid positions (0004). The specification further teaches that mutations resulting in IRT resistant are distinct from those leading to ESBL.

The specification further teaches that for any codon there are 64 permutations of nucleic acids (0030).

The specification teaches "The present invention also provides information about mutations having occurred in the beta-lactamase gene not yet know" (0033).

The specification further lists in Table 1, 88 codons of TEM beta lactamase (44 wildtype and 44 mutant by comparison to TEM-1).

The specification further teaches that 119 SNPs of the TEM beta lactamase were known at the time of filing (0051).

The specification further teaches 44 probes for known TEM beta lactamase (table 2). The specification teaches these probes represent most of the amino acid substitutions known at the time (0053).

The specification teaches probes of SEQ ID NO 5 to 45. It is noted that probes 5 to 45 have an N at the polymorphic position and thus encompass any nucleotide at that position. Thus the SEQ ID NO 5 to 45 will detect the presence of the sequences on either side of the N nucleotide, but will not predictably teach the genotype at the position of N.

The specification teaches in the example, "All the perfect match positions for blaTEM-1 could be identified correctly in every sample. The mean mismatch relative intensities vary, most of them remain below 0.4, only S143, S161, S182, S276 show higher values up to 0.6. The standard deviation varies from 0.01 to 0.21. For most positions the [(MM or PM)/PM] ratio is reproducible $\pm 10\%$. Only for Positions 127, 235, 236.2, 276 the variation is slightly higher, but the perfect match identification is without ambiguity" (0083). While the specification teaches hybridization of the sequence from the sample allowed identification without ambiguity of the perfect match sequences, it teaches that the mismatch sequences were not determined without ambiguity. As the specification teaches the mismatch sequences (mutant or antibiotic resistant) could not be identified without ambiguity it would not be able to predictably determine the particular polymorphism of the TEM beta-lactamase.

Further as the specification teaches that both the wildtype and mutant sequences hybridized to the array, it would be unpredictable to determine the presence of a beta-lactam resistant organism based on the hybridization.

The state of prior art and the predictability or unpredictability of the art:

Persson teaches hybridization of oligonucleotides is affected by length, temperature and complementarity (Analytical biochemistry (1997) 246 page 34-44) (see abstract). Persson teaches a fundamental problem in determining the sequence by hybridization is to resolve ambiguities at due to mismatches at the terminal ends of the hybrids (see page 34, 2nd column, 1st full paragraph). Persson further teaches that addition of a single nucleotide can increase stability of a complex more than 10 fold (see page 41, 2nd column). Thus Persson teaches that hybridization assays are not predictable at any probe length, temperature, percentage of complementarity, target length, or target concentration.

Draghici (Trends in Genetics (2006) volume 22, pages 101-109) teaches that increase the probe length from 25 to 30 nucleotides can increase sensitivity by 10 fold (see page 103, 1st column, 1st full paragraph) Draghici teaches that cross hybridization between probes that target the same region of a target sequence can result in unpredictable results (see page 107, 2nd column, last paragraph).

Kurkowa (Antimicrobial Agents and Chemotherapy (2003) volume 47, pages 2981-2983) teaches identification of a TEM-91 mutant that is resistant to ceftamizidime due to 3 mutations at codons 164, 184, and 240 (see abstract). Kurkow teaches that mutant has different kinetics than TEM 72 and TEM 5, TEM 46 and TEM 87 that have

either the same mutations or mutations in amino acids that are proximal to codons 164, 184, 240. Kurkow thus teaches one could not predictably infer antibiotic resistance based on the presence of a mutant.

Ning et al (Nature Genetics 1996 volume 14, pages 86-89 and SCORE results) teach that SEQ ID NO 7 of the instant specification has 100% identity with GenBank Accession Z96537 GI:2181612 which is a human subtelomeric probe. Thus Ning et al teaches that sequences of the instant claims cannot be predictably associated with beta-lactam resistant microorganism, as the sequences are present in humans, which are not microorganisms. Thus performing the claimed method using the probes taught by the specification would identify a sample taken from a human as containing a TEM beta-lactam resistant sample. It thus is unpredictable.

The level of skill in the art:

The level of skill in the art is deemed to be high.

Quantity of experimentation necessary:

In order to practice the invention as claimed, one would first have to establish that a predicative relationship exists between any nucleic acid sequence that comprises any of the codons recited in Table 1 and any 3 to 20 nucleotides to the 3' or 5' end codons, as all naturally occurring nucleotides specific to the TEM beta lactamase gene. This would be unpredictable as the claims broadly encompass every sequence in every genome, known or unknown. It would thus be unpredictable to associate hybridization to probes of any sequence with the presence of a beta lactam resistant antibiotic.

It would be unpredictable to associate the presence of hybridization to any probe sequences taught by SEQ ID NO 5 to SEQ ID NO 45 or the probes of claim 1 as Ning (in view of the SCORE alignment) demonstrates that SEQ ID NO 7 of the instant specification is identical to Accession Z96537 GI:2181612 which is a human subtelomeric probe. Thus hybridization of this probe to the array would not predictably allow identification of microorganisms that are beta-lactam resistant, as SEQ ID No 7 would identify humans as a beta-lactam resistant microorganism.

Further it would be unpredictable to determine the genotype based on the probes of SEQ ID No 5 to 45 as required in claim 15 SEQ ID NO 5 to 45 contain a N representing any nucleotide. It would be thus unpredictable to determine a genotype by using probe sequences that are degenerate or require a combination of all possible nucleotides as that position. Further claim 15 is unpredictable as it is drawn to determination of the resistance and genotype based on the hybridization to a specific position on the array, but does not require the probes be at predetermined position (as claim 1 requires). It thus would be unpredictable to determine a genotype without having a predetermined location for probes on the array.

Further it would be unpredictable to determine the presence of a particular polymorphism by hybridization as the specification teaches that mutant sequence could not be determined without ambiguity.

Further it would be unpredictable to identify the presence of beta-lactam resistant micro-organism with probes of different lengths, at different temperatures, with different

target nucleic concentrations as Draghici and Persson teach that all of these factors alter the reproducibility and thus predictability of microarrays.

Due to the scope of the claims, one of skill in the art would be required to further undertake extensive trial and error experimentation.

Therefore, in light of the breadth of the claims, the lack of guidance in the specification, the high level of unpredictability in the associated technology, the nature of the invention, the negative teachings in the art, and the quantity of unpredictable experimentation necessary to practice the claimed invention, it would require undue experimentation to practice the invention as claimed.

Response to Arguments

The response asserts that in view of the amendment the claims are enabled. These arguments have been thoroughly reviewed, but are not considered persuasive as Ning demonstrates that nucleic acids from humans in a biological sample would hybridize to the claim array (SEQ ID NO 7) and thus the claimed method would detect a human sample as as a beta-lactam resistant microorganism. It would thus be unpredictable to practice the invention as claimed to detect microorganisms in a biological sample as it would incorrectly identify a human as a microorganism that is beta-lactam resistant.

Claim Rejections - 35 USC § 103-maintained

4. The following is a quotation of 35 U.S.C. 103(a) which forms the basis for all obviousness rejections set forth in this Office action:

(a) A patent may not be obtained though the invention is not identically disclosed or described as set forth in section 102 of this title, if the differences between the subject matter sought to be patented and the prior art are such that the subject matter as a whole would have been obvious at the time the invention was made to a person having ordinary skill in the art to which said subject matter pertains. Patentability shall not be negated by the manner in which the invention was made.

This application currently names joint inventors. In considering patentability of the claims under 35 U.S.C. 103(a), the examiner presumes that the subject matter of the various claims was commonly owned at the time any inventions covered therein were made absent any evidence to the contrary. Applicant is advised of the obligation under 37 CFR 1.56 to point out the inventor and invention dates of each claim that was not commonly owned at the time a later invention was made in order for the examiner to consider the applicability of 35 U.S.C. 103(c) and potential 35 U.S.C. 102(e), (f) or (g) prior art under 35 U.S.C. 103(a).

5. Claims 1-6, 8, 9, 11, 13, and 15 are rejected under 35 U.S.C. 103(a) as being unpatentable over Lee, et al (Molecules and Cells (2002) volume 14, pages 192-197) in view of Blazquez, et al (Antimicrobial Agents and Chemotherapy (1995) volume 39, pages 145-149), Chee et al (A) (WO 95/11995), and Sutcliffe (Proceedings National Academy of Sciences USA (1978), volume 75 pages 3737-3741). This rejection is consistent with the scope of enablement as a biological sample encompasses the use of a bacterial sample.

Claim 1 is drawn to obtaining a biological sample, optionally isolating DNA from the sample and contacting the DNA from the sample with an array with capture probes derived from the sequence of a beta-lactamase genes to determine the presence of a beta lactamase resistant organism and mutations indicative of resistance to specific antibiotics.

With regards to claims 1, Lee teaches the use of arrays in which hybridization is indicative of beta-lactamase resistance (see abstract). Lee further teaches hybridization of target DNA with a DNA-chip, or micro-array, with probe sets at specific locations on the chip (see page 93 hybridization, preparation of DNA chip) to determine the presence of beta lactamase resistant genes (see figures 1, 3, 4) using probes of 21 nucleotides.

With regards to claims 2 and 3, Lee teaches isolation and amplification of target DNA from bacterial cells prior to contacting with array (see page 193, last paragraph 1st column and 1st paragraph 2nd column).

With regards to claims 8, Lee et al teaches a micro-array for the detection of various beta-lactamase resistant genes, including PSE, OXA, FOX, MEN, CMY, TEM, SHV, OXY, and *AmpC* (see abstract).

With regards to claim 11 and 13, Lee teaches the fluorescent labeling of DNA prior to contact with array (see page 193, column 1 last paragraph 3rd line up).

Lee et al does not teach a set of probes with all combinations of probes comprising R1- (X)-R2 of a beta lactamase gene (claim 1), fragmentation of DNA prior to contacting it with array (claim 4), beta-lactamases from Enterobacteriaceae (claim 5), or known SNPs in the beta-lactamase gene (claim 6).

However, with regards to claims 1,6, Blazquez et al teaches mutations, or SNPs, of beta-lactamase, including the mutation of gln39lys (specification refers to gln37lys) (see figure 1) alter beta lactamase stability and antibiotic activity (see page 148 column 1, 1st 4 lines of next to last paragraph). (It is noted that there are two accepted number systems for beta-lactamases, the Ambler numbering system based used by Blazquez

and the Sutcliffe system used in the specification. The two numbering systems may stem from Sutcliffe's sequencing pBR322 and Ambler sequencing the protein product of R6K (Proceedings National Academy of Sciences, USA (1978), Volume 75 pages 3732-3736). The art are normally presents mutants in reference to either Ambler or Sutcliffe numbering. The specification does not explicitly state a numbering system. Neither numbering system has glu at both codon 37 and 39. It is thus presumed codon 37 of specification and codon 39 of Blazquez are the same barring proof otherwise.)

Blazquez further teaches introduction of known mutations into the TEM1 gene at codons 39, 104, 164, 237, 238, and 240 alter resistance of microorganisms to specific antibiotics (see table 2 and 3). Mutation of amino acid 39 specifically decreases the susceptibility to cephaloridine and ceftazidime, but not amoxicillin, amoxicillin plus cavulanic acid, cefotaxime, aztreonam, and meropenem (see tables 2 and 3, and page 146 2nd column paragraph (i).

With regards to claim 5, Blazquez teaches E.coli which is a member of the Enterobacteriaceae family.

With regards to claims 1 and 4, Chee (A) et al teaches a tiling array (see Figure 7 and page 37 line 10- page 38 line 34). Chee (A) teaches the use of immobilized arrays to interrogate a reference sequence and its codons with a target sequence for the identification of single base mutants in the reference sequence associated with disease (see page 31 lines 6-7, and page 11 line 9 and 10). Further Chee (A) teaches this approach allows simultaneous detection and quantification of multiple target sequences (see page 32 lines 18-19), allowing for sequence determination. The block-tiling array

allows the interrogation of multiple nucleotide sites by use of multiple probe sets, which represent every permutation of nucleotides possible for a given nucleotide sequence.

Chee (A) teaches the determination of all possible combinations of nucleotides surrounding a SNP using from 15-30 nucleotides (page 27 lines 2-6), allowing determination of all possible nucleic acid sequences. With regards to claim 4, Chee et al teaches DNA fragmentation (see page 126, number 4), prior to contacting with capture probes. Chee (A) teaches microfabricated arrays with large numbers of oligonucleotides offer great promise (see page 2 lines 11-13) for applications including identification of mutations related to disease, forensic studies, epidemiological and forensic studies. Chee (A) further teaches, "It is desirable to simultaneously diagnose the presence or absence of a variety of lethal common infections, determine the most effective therapeutic regime" (see page 64 lines 25-28). Chee teaches, "The length of a reference sequence can vary widely from a full-length genome, to an individual chromosome, episome, gene" (page 20, lines 36-38).

With regards to claims 1 and 6, Sutcliffe teaches the nucleotide sequence of the beta- lactamase gene (see figure 3).

Therefore, it would be prima facie obvious to one of ordinary skill in the art at the time the invention was made to improve the method of identifying beta-lactamase resistance taught by Lee, to include identification of specific mutations resulting in resistance to specific antibiotics as taught by Blazquez with reasonable expectation of success. The ordinary artisan would be motivated to combine the teaching of Lee and Blazquez in order to identify the specific beta lactam resistance present and provide

proper treatment. It would have been further prima facie obvious to the ordinary artisan at the time the invention was made to improve the method of identifying specific mutations for specific resistance of Lee and Blazquez to incorporate a tiling array as taught by Chee (A) and use the beta lactamase sequence taught by Sutcliffe to allow simultaneous detection and quantification of multiple target sequences (see page 32 lines 18-19), thus allowing thorough characterization of mutations known to alter beta lactam resistance, and possibly identify new mutations altering beta-lactam resistance . The ordinary artisan would be motivated to improve the method of Lee and Blazquez with the use of a tiling array as taught by Chee (A), because Chee (A) teaches simultaneously diagnose the presence or absence of a variety of lethal common infections, determine the most effective therapeutic regime (see page 64 lines 25-28). In performing the method of Lee, Blazquez, Chee (A), and Sutcliffe, the ordinary artisan would be motivated to specifically include a probe in the screening method, corresponding to the mutation at codon 37 (same as codon 39 taught by Blazquez), because Blazquez teaches that a mutation at this codon results in specific antibiotic resistance. The ordinary artisan would be motivated combine the teachings of Lee, Blazquez, Chee (A), and Sutcliffe to screen for the presence of mutations at codon 37 (codon 39 taught by Blazquez) with the use of tiling probes as taught by Chee (A) to detect the susceptibility to cephaloridine and ceftazidime. The ordinary artisan would therefore be motivated to construct a probe corresponding to mutations in codon 37, including a probe with the sequence of SEQ ID NO: 7 (claim 9), to provide tiling probes with all possible combinations of nucleotides at codon 37. The artisan would have had

a reasonable expectation of success using the known methods of array based nucleic acid hybridization, to detect known mutations by probes designed by known methods.

It would have further been prima facie obvious to one of ordinary skill in the art at the time the invention was made to combine the teachings of Lee, Blazquez, Chee and Sutcliffe to use a microarray comprising probes that are functionally equivalent to the probes claimed by SEQ ID NO 5 to 45. The artisan would be motivated to make and use such an array in because Chee teaches that tiling arrays for scanning of entire genes for mutations and Lee and Blazquez teach that TEM gene is an important gene in beta-lactamase resistance. The artisan would have a reasonable expectation of success as the sequence of beta-lactamase is known and Chee teaches methods of making arrays.

Response to Arguments

The response asserts the present method is drawn to a microarray with 44 triplets provided in table 1 and their mutant counter part. This argument has been thoroughly reviewed but is not considered persuasive as the claim 1 requires a microarray comprising different sets of capture probes, where each set consists of a nucleotide triplet identified in table 1 as either a wildtype or mutant sequences. Thus the contrary to the assertion of the response the claims do not require the 44 wildtype and 44 mutant triplets of table 1, but merely different probe sets that are identified as wildtype or mutant in table 1.

The response then briefly reviews the teachings of the art on the top of page 9.

The response asserts that the cited materials do not teach or suggest the use of capture probes utilizing the 44 triplets provided in table 1. These arguments have been thoroughly reviewed but are not considered persuasive as claim 1 does not require the 44 probes of table 1, but different sets of probes comprising a wildtype or mutant capture probes.

The response further asserts that the teachings do not teach or suggest a microarray comprising SEQ ID NO 5 to 45 of claim 15. This argument has been thoroughly reviewed but is not considered persuasive as the teachings of Lee, Blazquez and Sutcliffe teach the sequence of the bla-TEM beta lactamase and its importance in beta-lactamase resistance and Chee teaches a method of making a tiling array for determining the sequence of an entire gene. Thus as outlined it would have been prima facie obvious to one of ordinary skill in the art at the time the invention was made to combine the teachings of Lee, Blazquez, Sutcliffe, and Chee to make an array that comprises probes that are functionally equivalent to those in SEQ ID NO 5 to 45 and use them in a method to identify beta lactamase resistance.

The response further asserts that the examiner has not provided motivation to combine the elements of the instant invention and a reasonable expectation of success. These arguments have been thoroughly reviewed but are not considered persuasive as KSR forecloses the argument that a specific teaching, suggestion or motivation is required to support obviousness. Further the examiner stated in the previous action, "The ordinary artisan would be motivated combine the teachings of Lee, Blazquez, Chee (A), and Sutcliffe to screen for the presence of mutations at codon 37 (codon 39

taught by Blazquez) with the use of tiling probes as taught by Chee (A) to detect the susceptibility to cephaloridine and ceftazidime. The ordinary artisan would therefore be motivated to construct a probe corresponding to mutations in codon 37, including a probe with the sequence of SEQ ID NO: 7 (claim 9), to provide tiling probes with all possible combinations of nucleotides at codon 37. The artisan would have had a reasonable expectation of success using the known methods of array based nucleic acid hybridization, to detect known mutations by probes designed by known methods." Thus the examiner has provided motivation and reasonable expectation of success.

The response further asserts there is no teaching contemplates the detection and genotyping of the genotype in a method that specifically employs the claimed arrays. This argument has been thoroughly reviewed but is not considered persuasive as the detection and sequencing by the combination of the teachings of Lee, Blazquez, Chee and Sutcliffe would result in the detection of the presence or absence of the wildtype or mutant sequence of the bla-TEM, thus rendering the instant claims obvious as it would detect and genotype simultaneously.

6. Claim 7 is rejected under 35 U.S.C. 103(a) as being unpatentable over Lee, Blazquez, Chee (A) and Sutcliffe, as applied to claims 1-6, 8, 9, 11, and 13 above, in further view of Osano et al (Antimicrobial Agents and Chemotherapy (1994) volume 38, pages 71-78)

Claim 7 further limits the enterobacteriaceae beta lactamase of claim 5 to a serine or zinc beta-lactamase.

The teachings of Lee, Blazquez, Chee (A) and Sutcliffe are set forth above in paragraph 8. Lee, Blazquez, Chee (A), and Sutcliffe, do not teach on serine or zinc beta-lactamase from the bacterial family Enterobacteriaceae.

However, Osana et al, teaches *S. marcescens* is a member of enterobacteriaceae family (see page 76 column 1, lines 16-19). Osana further teaches class A and C beta-lactamases, including plasmid encoded TEM and SHV, are serine dependent (see page 71, column 1, lines 5 and 6 column 1 and column 2 lines 4-7). Osana further teaches class B beta-lactamases are zinc dependent (see page 71 column 2 lines 4-7). Osana teaches that some strains of the Enterobacteriaceae family are reported to be resistant to imipenem therapy. Osana teaches IMP-1 a zinc beta lactamase confers imipenem resistance to *S. marcescens* and further demonstrates there is great variability in the amino acid sequence of known zinc beta-lactamases and suggests evolution independent of other known zinc beta lactamases (see figure 3).

Therefore it would have been prima facie obvious to the ordinary artisan at the time of the invention was made, to improve the method taught by Lee, Blazquez, Chee (A) and Sutcliffe, to include the zinc beta-lactamases as taught by Osano, et al for the purpose of detecting imipenem resistance in the Enterobacteriaceae family. The artisan would have a reasonable expectation of success as the combination would merely be using methods known to detect mutations in the prior art to detect known mutations in known nucleic acid sequences. The ordinary artisan would be motivated to use the method taught by Lee, Blazquez, Chee (A), and Sutcliffe to determine the presence of

zinc beta lactamases in order to identify imipenem resistant strains, to allow for proper diagnosis and treatment.

Response to arguments

Applicant asserts reasons discussed above that Chee, Sutcliffe, Lee and Blazquez do not meet the limitations of the independent claims and thus do not cure the defects of claim 7.

Applicant's arguments have been fully considered by they are not persuasive. The response asserts Claim 7 depends from Claim 5 and ultimately Claim 1. The response states that Lee, Blazquez and Chee (A) are discussed above and Osono does not cure the defects of the primary or secondary references. This argument is not persuasive for the reasons presented above for Lee in view of Blazquez and Chee (A) and Sutcliffe. Lee, Blazquez, Chee , and Sutcliffe combine to teach each and every limitation of the claims from which claim 7 depends, further Osano teaches detection of zinc beta lactamases, thus Osana renders all of the the limitations of claim 7 obvious.

Claim 10 is rejected under 35 U.S.C. 103(a) as being unpatentable over Lee, Blazquez, Chee (A), and Sutcliffe as applied to claims 1-6, 8, 9, 11, and 13, further in view of Chee (B) et al (Science (1996), Volume 274, pages 610-614) and Routier (Nucleic Acids Research, (1999) volume 27, pages 4160-4166).

The teachings of Lee, Blazquez, Chee (A) and Sutcliffe are set forth above. Lee, Blazquez, Chee (A), and Sutcliffe do not teach fragmentation of DNA to 15-50 nucleotides.

However, Chee (B) teaches fragmentation improves the uniformity and specificity of hybridization (see page 613 third column, lines 43 and 44). Routier teaches a method of fragmentation resulting in fragments of 15-50 nucleotides (see Figure 5).

Therefore it would be prima facie obvious for one of ordinary skill the art at the time of the invention to modify the method of Lee, Blazquez, Chee (A), and Sutcliffe for detection of beta lactamase resistance with the Routier method of DNA fragmentation wherein the fragments are 15-50 nucleotides. Routier teaches fragmentation with sizes of 15-50 nucleotides and Chee (B) teaches fragmentation improves uniformity and specificity of hybridization. The ordinary artisan would be motivated to optimize the size of fragments of the DNA prior to contacting with a microarray because Chee (B) teaches it improves specificity and uniformity of hybridization. The artisan would have a reasonable expectation of success as Chee (B) teaches the use of fragmentation for microarray analysis as taught by the combination of. Lee, Blazquez, Chee (A), and Sutcliffe.

As stated in the MPEP, 2144.05 II, "[W]here the general conditions of a claim are disclosed in the prior art, it is not inventive to discover the optimum or workable ranges by routine experimentation." In re Aller, 220 F.2d 454, 456, 105 USPQ 233, 235 (CCPA 1955)."

Response to arguments

Applicant asserts reasons discussed above that Chee, Sutcliffe, Lee and Blazquez do not meet the limitations of the independent claims and thus do not cure the defects of claim 10.

7. Applicant's arguments have been fully considered by they are not persuasive. The response asserts Claim 10 depends from Claim 4 and ultimately Claim 1. The response states that Lee, Blazquez and Chee (A) are discussed above and Chee (B) and Routier do not cure the defects of the primary or secondary references. This argument is not persuasive for the reasons presented above for Lee in view of Blazquez and Chee (A) and Sutcliffe. As Lee, Blazquez and Chee render the claims from which claim 10 depends and Routier teaches fragmentation, the instant rejection renders claim 10 obvious, as well.

8. Claim 12 is rejected under 35 U.S.C. 103(a) as being unpatentable over Lee, Blazquez, Chee (A), and Sutcliffe as applied to claims 1-6, 8, 9, 11, and 13 above, and further in view of Behrendorf, et al (Nucleic Acids Research (2002) volume 30, e64).

The teachings of Lee, Blazquez, Chee (A) and Sutcliffe are set forth above. Lee, Blazquez, Chee (A) and Sutcliffe do not teach labeling of DNA following contacting DNA with array.

However, Behrendorf, et al teaches the detection of SNPs by binding fluorescently labeled mutS to mismatched DNA for detection of SNPs (see figure 1) on an array. Behrendorf teaches this method allows, "robust detection of genetic variation," while decreasing hybridization times and shortening assay duration (see page 5, 1st column lines 1 and 2, and 2nd column lines 10-12).

Therefore it would be prima facie obvious to the ordinary artisan at the time the invention was made to improve the array based method of detecting the presence of beta-lactam resistant bacteria of Lee, Blazquez, Chee (A) and Sutcliffe by using fluorescently labeled mutS taught by Behrendorf to label the DNA after contacting with array, because Behrendorf teaches that it gives robust detection of genetic variations, decreases hybridization times and shorten assay duration. The ordinary artisan would be motivated to use the labeling method of Behrendorf in the method of Lee, Blazquez, Chee (A) and Sutcliffe because Behrendorf teaches that it gives robust detection of genetic variations, decreases hybridization times and shorten assay duration. The artisan would have a reasonable expectation of success as combining known methods for detecting nucleic acid mutations in an art accepted manner.

Response to arguments

9. Applicant asserts for reasons discussed above that Chee, Sutcliffe, Lee and Blazquez do not meet the limitations of the independent claims and thus do not cure the defects of claim 12.

Applicant's arguments have been fully considered by they are not persuasive. The response asserts Claim 12 depends from Claim 1. The response states that Lee,

Blazquez and Chee (A) are discussed above and Behrendorf does not cure the defects of the primary or secondary references. This argument is not persuasive for the reasons presented above for Lee in view of Blazquez and Chee (A) and Sutcliffe. The teachings of Lee, Blazquez and Chee render the claim 1 obvious, thus as Behrendorf teaches labeling after the contacting the sample nucleic acids to an array, the claim 12 is also obvious.

Summary

No claims are allowed over prior art cited.

Conclusion

10. Applicant's amendment necessitated the new ground(s) of rejection presented in this Office action. Accordingly, **THIS ACTION IS MADE FINAL**. See MPEP § 706.07(a). Applicant is reminded of the extension of time policy as set forth in 37 CFR 1.136(a).

A shortened statutory period for reply to this final action is set to expire **THREE MONTHS** from the mailing date of this action. In the event a first reply is filed within **TWO MONTHS** of the mailing date of this final action and the advisory action is not mailed until after the end of the **THREE-MONTH** shortened statutory period, then the shortened statutory period will expire on the date the advisory action is mailed, and any extension fee pursuant to 37 CFR 1.136(a) will be calculated from the mailing date of the advisory action. In no event, however, will the statutory period for reply expire later than **SIX MONTHS** from the date of this final action.

Any inquiry concerning this communication or earlier communications from the examiner should be directed to Steven C. Pohnert whose telephone number is (571)272-3803. The examiner can normally be reached on Monday-Friday 6:30-4:00, every second Friday off.

If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Ram Shukla can be reached on 571-272-0735. The fax phone number for the organization where this application or proceeding is assigned is 571-273-8300.

Information regarding the status of an application may be obtained from the Patent Application Information Retrieval (PAIR) system. Status information for published applications may be obtained from either Private PAIR or Public PAIR. Status information for unpublished applications is available through Private PAIR only. For more information about the PAIR system, see <http://pair-direct.uspto.gov>. Should you have questions on access to the Private PAIR system, contact the Electronic Business Center (EBC) at 866-217-9197 (toll-free). If you would like assistance from a USPTO Customer Service Representative or access to the automated information system, call 800-786-9199 (IN USA OR CANADA) or 571-272-1000.

Steven Pohnert

/Sarae Bausch/
Primary Examiner, Art Unit 1634